



## RESEARCH ARTICLE

# Deep learning-based fully automatic segmentation of wrist cartilage in MR images

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The study objective was to investigate the performance of a dedicated convolutional neural network (CNN) optimized for wrist cartilage segmentation from 2D MR images. CNN utilized a planar architecture and patch-based (PB) training approach that ensured optimal performance in the presence of a limited amount of training data. The CNN was trained and validated in 20 multi-slice MRI datasets acquired with two different coils in 11 subjects (healthy volunteers and patients). The validation included a comparison with the alternative state-of-the-art CNN methods for the segmentation of joints from MR images and the ground-truth manual segmentation. When trained on the limited training data, the CNN outperformed significantly image-based and PB-U-Net networks. Our PB-CNN also demonstrated a good agreement with manual segmentation (Sørensen–Dice similarity coefficient [DSC] = 0.81) in the representative (central coronal) slices with a large amount of cartilage tissue. Reduced performance of the network for slices with a very limited amount of cartilage tissue suggests the need for fully 3D convolutional networks to provide uniform performance across the joint. The study also assessed inter- and intra-observer variability of the manual wrist cartilage segmentation (DSC = 0.78–0.88 and 0.9, respectively). The proposed deep learning-based segmentation of the wrist cartilage from MRI could facilitate research of novel imaging markers of wrist osteoarthritis to characterize its progression and response to therapy.

## KEYWORDS

applications, cartilage, human study, methods and engineering, musculoskeletal, postacquisition processing, quantitation

## 1 | INTRODUCTION

MRI is a versatile tool for the detection of morphological and compositional cartilage abnormalities in degenerative diseases of joints.<sup>1</sup> MRI-based measurements of a joint space narrowing<sup>2-4</sup> have been utilized to assess cartilage degradation in multiple locations including knee<sup>5</sup> and wrist.<sup>6</sup> MRI has also been applied to quantify other morphometric features including cartilage cross-sectional area (CSA)<sup>7</sup> and cartilage volume.<sup>8,9</sup> More recently, several quantitative MRI approaches have been proposed to assay proteoglycan and collagen components of the cartilage matrix.<sup>10-13</sup> It has been suggested that such biomarkers could be used for disease detection and treatment monitoring.

Analysis of structural and quantitative MRI data requires an accurate cartilage segmentation, whose automation for routine applications is challenged by the presence of other tissues with similar MR contrast (eg, muscles, skin, edematous tissues). On that basis, a manual segmentation is considered the gold standard in cartilage assessment applications.<sup>8,14-17</sup> However, manual segmentation is a highly time-consuming and tedious task and its reliability can be hampered by inter-operator variability. To improve the speed and consistency of the cartilage segmentation, a wide variety of computer-assisted approaches have been proposed, including semi-automated<sup>18,19</sup> and fully automatic<sup>20-23</sup> segmentation methods. These approaches provide very fast and reliable segmentation with a moderate penalty on the segmentation accuracy, ie, with a Sørensen–Dice similarity coefficient (DSC)<sup>24</sup> approaching 0.80.<sup>20</sup> Most recently, convolutional neural networks (CNNs)<sup>25-28</sup> have been successfully applied for the segmentation of knee MR images. The methods demonstrated improved DSC values of 0.82<sup>25</sup> and 0.88<sup>26</sup> for planar and U-Net<sup>29</sup> architectures, respectively. This highlighted initial promise of the machine learning approaches for the fully automated segmentation of complex anatomical structures.

MRI has proven to be a promising approach for wrist osteoarthritis (OA) evaluation, demonstrating higher sensitivity to moderate changes of OA compared with X-ray-based assessment.<sup>30</sup> MRI-based wrist joint assessment is potentially more suitable to follow changes over time and/or to assess the efficiency of therapy than CT arthrography given that harmful ionizing radiation and injection of a contrast material into the joint space are a part of the CT imaging procedure.<sup>31</sup> However, while most automated segmentation methods have been developed for knee MRI, only a few<sup>7,9</sup> techniques have been optimized for wrist MR image segmentation, likely due to the more complex anatomy of the wrist joint. The automatic segmentation of the wrist joint cartilage from MRI images could facilitate research of novel imaging biomarkers of wrist OA and to characterize its progression and response to therapy.

Given the outstanding performance of CNN-based approaches for the segmentation of knee structures,<sup>25,26</sup> we hypothesized that deep-layered CNNs could be valuable in the design of an automatic segmentation of wrist cartilage. Therefore, in the present work, we developed, optimized and evaluated a CNN-based method for a fully automatic segmentation of wrist joint cartilage. The network was trained using the manual labels produced by an experienced radiologist and compared with several representative CNN-based methods.<sup>26,32</sup>

## 2 | PATIENTS AND METHODS

### 2.1 | Subjects

The study was approved by the local ethics committee of Federal Almazov North-West Medical Research Center. Eleven subjects were enrolled into the study once their written informed consent had been obtained. The subjects consisted of eight healthy volunteers (no previous wrist trauma, six males and two females, aged 23-38 years, mean 29.6 years) and three patients (two females [aged 63 and 77 years] with a confirmed OA diagnosis, and one 62-year-old female with articular pain). All data were acquired in the dominant wrist.

### 2.2 | MR imaging

MR images were acquired with a 1.5 T Magnetom Espree system (Siemens GmbH, Erlangen, Germany). The same wrist was scanned twice, first with a conventional “birdcage”-type transmit/receive extremity coil and then with a homemade wireless coil providing a higher signal-to-noise ratio (SNR).<sup>33</sup> In two subjects, one of the scans was not completed due to technical or cooperation reasons, thereby bringing the total number of MRI scans to 20. 3D coronal T1-weighted gradient echo (Volumetric Interpolated Breath-hold Examination [VIBE]) images with water-selective excitation for fat suppression were acquired to achieve an optimized contrast-to-noise ratio for the cartilage.<sup>7</sup> The relevant parameters were: TR/TE = 18.6/7.3 ms, flip angle = 10°, FOV = 97 x 120 mm<sup>2</sup>, matrix size = 260 x 320, voxel size = 0.37 x 0.37 x 0.5 mm<sup>3</sup>, number of coronal slices = 88. The total acquisition time was 6 minutes.

### 2.3 | Data preparation

3D regions of interest (ROIs) were manually outlined to encompass all cartilage tissues in the acquired volumes. Every other slice was selected from all 3D ROIs to form an intermediate dataset containing 420 images with cartilage. Then the dataset was augmented by an additional set of 140 slices that did not contain cartilage tissue, giving a final dataset comprising 560 images.

### 2.3.1 | Dataset labeling

Image processing was performed using MATLAB (MathWorks, Natick, MA, USA). Cartilage tissue was segmented by an expert radiologist (O1: V.A.F.) using a software-assisted manual approach. In detail, the wrist joint ROI delineated for each slice by manual contouring (Figure 1A) was first roughly segmented by intensity thresholding, with threshold value optimized by the observer in iterative fashion on a per slice basis. Next, the resulting binary masks (Figure 1B) were manually corrected to ensure that only cartilage pixels were included in the labels.

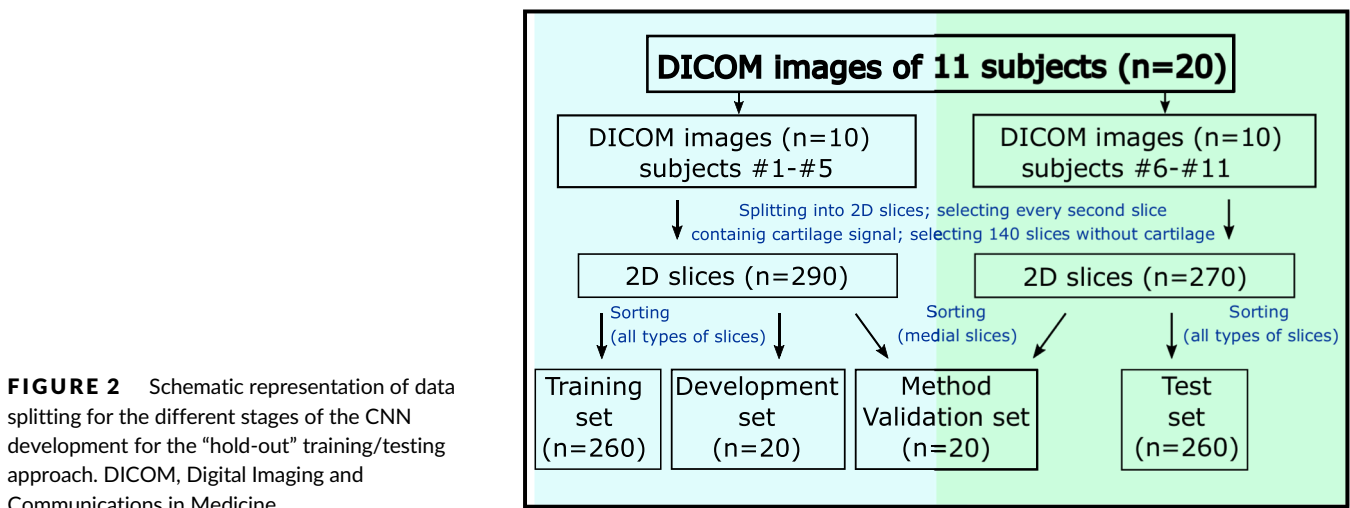
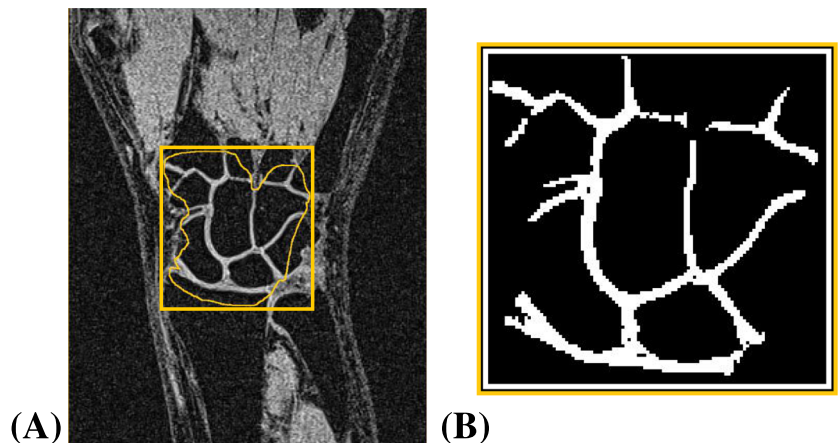
### 2.3.2 | Data splitting and CNN training approaches

The labeled images were split into several subsets to be used for the training ( $n = 260$ ), development ( $n = 20$ ), testing ( $n = 260$ ) and method validation ( $n = 20$ ) stages (Figure 2). We utilized a “hold-out” training approach, in which training and development datasets included data from healthy volunteers #1 to #5 (MRI scans #1 to #10) and the test dataset included data from subjects #6 to #11 (MRI scans #11 to #20). For each subject, the medial coronal slice chosen from the 3D dataset was used for the validation of our method and for comparison with the manual one. To ensure these images were unseen by the CNN, these slices were excluded from the training, development and test datasets.

To assess the CNN robustness with respect to the anatomical heterogeneity of the training dataset, several CNN variants were trained using different training subsets containing: (1) 5%, 10%, 20%, 33%, 50% and 66% of the total number of slices ( $n = 260$ ) randomly selected from the training dataset; and (2) images of particular subjects, starting from subject #1 and then subsequently adding other subjects' data.

Additionally, we performed cross-validation studies for three patients and three healthy volunteers included in the testing dataset to estimate the best achievable network performance in the presence of a limited number of subjects. In these studies, the network was trained using six different samples of 10 subjects (6-fold analysis) and tested on the remaining subjects (from #6 to #11).

**FIGURE 1** Illustration of the manual segmentation results. (A) Preliminary delineation of the wrist joint area. (B) Final binary mask obtained after thresholding and manual correction



For each CNN instance, the performance was evaluated by comparing the CNN-based and the manual segmentations using DSC values for 3D images or for planar slices independently. The SNR was measured as the ratio between the cartilage tissue signal and the standard deviation of the noise within a signal-free area.

## 2.4 | CNN-assisted segmentation

### 2.4.1 | Patch-based CNN architecture

The design of our network was based on our preliminary experiments, which indicated that the state-of-the-art U-Net-based CNN<sup>26</sup> did not perform satisfactorily, likely as a result of the small size of our training dataset (260 images) and the large number of trainable parameters ( $2.8 \times 10^6$ ). On that basis, and to minimize the risk of overfitting, we selected a planar network architecture with a smaller number of trainable parameters and a patch-based (PB) training approach, which proved to be adequate for a limited amount of annotated data.<sup>32,34,35</sup> The network parameters, including the number of convolutional layers, the number and the size of filters, were optimized by the grid search. The final PB-CNN architecture (Figures 3 and A1) had five convolutional layers with 44 filters of size  $3 \times 3$ . The patch size was further optimized for this architecture. Gaussian noise and drop-out regularization layers were added to reduce the generalization error and to minimize the risk of overfitting. The network was trained during 20 epochs with a 20 000 batch size. Ten percent of training data were used for the training validation (ie, for the calculation of loss value). The stopping criterion during the training phase was defined as the absence of loss value decrease for five epochs. A RMSProp optimizer (<https://keras.io/optimizers/>) with a default learning rate value of 0.001 was used.

### 2.4.2 | PB-CNN input and output

We used a sliding-window approach<sup>25</sup> to select patches surrounding the pixels to be classified. In detail, for each pixel of interest, a  $28 \times 28$  patch centered on the pixel was applied to provide a network input. For each image, the network output a probability map, which was thresholded to obtain cartilage binary masks. The threshold value was optimized during the development stage to maximize the DSC coefficient with respect to the development dataset. Overall, datasets for method development and validation contained  $1.3 \times 10^6$  patches. For the “hold-out” approach, both training and testing datasets contained a total of  $17.6 \times 10^6$  patches.

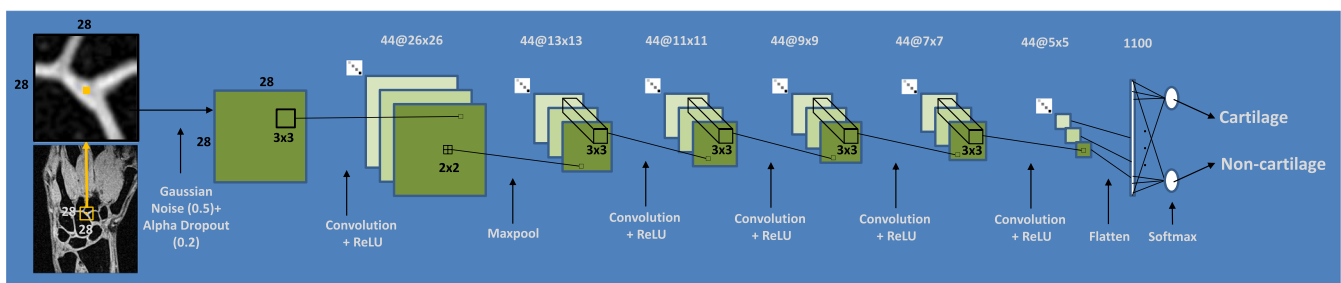
For the subject-based cross-validation, the training and testing samples were varied for every step of the 6-fold analysis. The number of patches in the training dataset varied from  $15.9 \times 10^6$  to  $16.7 \times 10^6$ , depending on how many times (one or two) each subject had been scanned. The networks were tested on one or two separate 3D images ( $1.8 \times 10^5$  patches) of subjects who were not included in the trained dataset.

### 2.4.3 | State-of-the-art neural networks

We compared our proposed network with several alternative architectures such as image-based<sup>26</sup> and PB<sup>32</sup>-U-Net CNNs trained and tested with the “hold-out” approach. These CNNs are detailed in the Appendix (Figures A2 and A3).

### 2.4.4 | Hardware and software

The training was performed on a server with four processors (Intel Xeon E5-4617 2.90 GHz) and 512 Gb RAM. To provide realistic estimates of the network execution speed, all methods were tested on a PC with more common characteristics (an Intel Core



**FIGURE 3** Configuration of PB-CNN optimized for wrist cartilage segmentation

i5-7640X processor with 32 Gb of RAM). CNNs were built using Python 3.6.4, TensorFlow 1.7.0, and Keras 2.1.5 open-source neural network library.

## 2.5 | Data analysis

### 2.5.1 | Reproducibility of the manual segmentation procedure

Reproducibility of the manual segmentation procedure used to create labels from the training dataset was assessed from the segmentation results for the method validation dataset obtained by three observers, each with more than 10 years of experience in musculoskeletal segmentation. The first observer (O1: V.A.F.) segmented the wrist cartilage twice. The segmentation sessions were separated by 1 week. The segmentation results from the first session were considered as the ground truth for the purposes of the method evaluation. Two other experts (O2: A.Y.E. and O3: R.F.) segmented images once.

Cartilage CSA was calculated as a product of the number of pixels within the binary mask and the pixel area ( $0.37 \times 0.37 \text{ mm}^2$ ). For each observer, the averaged cartilage CSA was calculated among the method validation dataset and compared statistically using Student's t-tests. To determine the variability of the manual segmentation procedure, the inter- and intra-observer DSC =  $2|X \cap Y| / (|X| + |Y|)^{2.4}$  values were calculated, where X is a binary mask segmented by O1 in the first session, and Y is segmented either by O1 in the second session (intra-observer study) or by O2 and O3 (inter-observer study).

### 2.5.2 | PB-CNN-based cartilage segmentations

Performances of the trained networks were evaluated based on comparisons with the ground truth, ie, the manual segmentation as described above. For the "hold-out" approach, the DSC value was calculated independently for each 3D volume from the test dataset and then averaged. The layer-to-layer analysis of the segmentation accuracy for the developed PB-CNN was performed according to two stages. Four cartilage zones along the slice selection direction in each 3D image were initially identified. Then the cartilage volume in each zone was normalized with respect to the volume of the medial slice in the corresponding 3D image, ie, the image containing the largest amount of cartilage, which was assumed to be 100%. The identification of the cartilage zones was designed to account for differences in wrist joint thickness among different volunteers. Zone #1 encompassed slices with no cartilage, zone #2 encompassed slices with a relative amount of cartilage up to 33%, zone#3 encompassed slices with an intermediate amount of cartilage from 34% to 66%, and zone#4 encompassed slices with an amount of cartilage ranging from 67% to 100%. An averaged DSC index was calculated for each zone of the 3D images.

In the cross-validation analysis, the trained networks were tested on 3D images (one or two) of the subjects not included in the training dataset. Corresponding DSC values were calculated for each 3D image as a whole and on the layer-to-layer basis.

The evaluation of the network trained using the "hold-out" approach was performed as follows: DSC was calculated individually for 20 medial coronal slices from the method validation dataset and compared with the human inter- and intra-observer study results. A radiologist with more than 10 years of experience in musculoskeletal MRI (O4: A.G.L.) performed a 10-point visual evaluation of the PB-CNN-based segmentations. Several factors of a segmentation quality were assessed; their presence in the cartilage mask led to a reduction of the initial score of 10 by:

3 points:

- segmentation of bone tissue pixels
- segmentation of pixels of pathological zones in bones
- segmentation of pixels out of the wrist joint;

2 points:

- a significant amount of nonsegmented cartilage tissue on articular surfaces of wrist joint;

and 1 point:

- nonsegmented cartilage tissue in three or more bones articulation
- deviations of the thickness of segmented cartilage
- segmented noncartilage pixels in wrist joint.

### 3 | RESULTS

#### 3.1 | Manual segmentation procedure

The averaged cartilage CSA determined by the first observer and considered as the ground truth was  $237.6 \pm 39.8 \text{ mm}^3$ . The corresponding result obtained by O2 ( $240.1 \pm 38.1 \text{ mm}^3$ ) was not significantly different ( $P > .05$ ), whereas O3 reported a significantly higher CSA ( $269.3 \pm 39.1 \text{ mm}^3$ ,  $P = .015$ ).

The average rate for the manual segmentation was 20 slices/hour (ie, 5 minutes per slice). Intra-observer DSC was  $0.90 \pm 0.04$  whereas inter-observer DSC values were lower ( $0.88 \pm 0.04$  for O1 vs. O2 and  $0.78 \pm 0.06$  for O1 vs. O3). The corresponding statistics for the manually performed segmentation procedure are summarized in Table 1. While the SNR of images differed from one subject to another due to the differences related to coil load, the SNR was systematically higher ( $P > .05$ ) with the homemade coil. The paired t-test ( $P > .05$ ) for the data of subjects scanned twice did not provide any significant difference in the segmentation accuracy for either intra- or inter-observer studies.

#### 3.2 | CNN-assisted segmentation

##### 3.2.1 | Performances of CNNs and sensitivity analysis

Figure 4 shows the results of the sensitivity analysis from the "hold-out" training/testing stage. The DSC value for the development dataset increased continuously up to DSC = 0.86 when the data of each subject were subsequently added to the training dataset. When the slices for

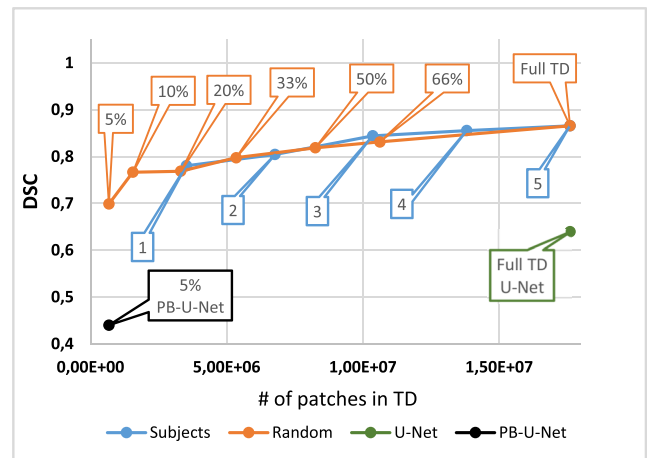
**TABLE 1** Statistics for manual and PB-CNN-assisted segmentation procedures. Quantitative and qualitative validation of methods

# of scan	SNR	Intraobserver DSC	Interobserver DSC (O1 – O2)	Interobserver DSC (O1 – O3)	Intermethod DSC (O1 – CNN)	Visual evaluation score for CNN
1	14.0	0.91	0.92	0.83	0.83	8
2	19.0	0.94	0.84	0.75	0.81	7
3	12.4	0.89	0.90	0.83	0.83	7
4	15.7	0.90	0.86	0.73	0.77	4
5	14.5	0.85	0.83	0.75	0.84	7
6	19.6	0.92	0.81	0.75	0.82	7
7	13.8	0.87	0.92	0.81	0.88	9
8	20.8	0.93	0.88	0.82	0.84	7
9	13.7	0.97	0.91	0.81	0.86	8
10	18.9	0.98	0.87	0.77	0.83	9
11	12.0	0.82	0.89	0.66	0.82	7
12	14.2	0.96	0.89	0.74	0.84	9
13	20.6	0.89	0.89	0.83	0.81	5
14	10.2	0.91	0.86	0.64	0.74	5
15	15.1	0.87	0.94	0.85	0.81	6
16	13.4	0.90	0.91	0.79	0.79	6
17	21.0	0.83	0.89	0.82	0.75	5
18	18.6	0.93	0.92	0.83	0.80	4
19	12.5	0.86	0.81	0.83	0.77	5
20	15.4	0.96	0.82	0.72	0.69	3
HV group Mean $\pm$ SD	<b>15.57 <math>\pm</math> 3.37</b>	<b>0.91 <math>\pm</math> 0.04</b>	<b>0.88 <math>\pm</math> 0.03</b>	<b>0.77 <math>\pm</math> 0.06</b>	<b>0.82 <math>\pm</math> 0.03</b>	<b>7.00 <math>\pm</math> 1.51</b>
P group Mean $\pm$ SD	<b>16.18 <math>\pm</math> 3.57</b>	<b>0.90 <math>\pm</math> 0.05</b>	<b>0.87 <math>\pm</math> 0.05</b>	<b>0.80 <math>\pm</math> 0.04</b>	<b>0.76 <math>\pm</math> 0.05</b>	<b>4.60 <math>\pm</math> 1.14</b>
All Groups Mean $\pm$ SD	<b>15.7 <math>\pm</math> 3.34</b>	<b>0.90 <math>\pm</math> 0.04</b>	<b>0.88 <math>\pm</math> 0.04</b>	<b>0.78 <math>\pm</math> 0.06</b>	<b>0.81 <math>\pm</math> 0.05</b>	<b>6.40 <math>\pm</math> 1.76</b>

Abbreviations: CNN, convolutional neural network; DSC, Sørensen–Dice similarity coefficient; HV, healthy volunteers (MRI scans #1–#15); O1, observer 1; O2, observer 2; O3, observer 3; O4, observer 4; P, patients (MRI scans #16–#20); SD, standard deviation; SNR, signal-to-noise ratio.

Bold font refers to the averaged data

**FIGURE 4** Dependence of DSC value on the training data amount and sample selection (“hold-out” training/testing approach). Blue dots correspond to consecutive inclusion of the data of each subject (from #1 to #5) to the training dataset (TD). Orange dots correspond to a random selection of slices for training in the indicated proportions from the whole TD (full TD). PB-U-Net, patch-based U-Net



training were randomly selected from the whole dataset in different proportions, DSC increased in a similar manner. The DSC improvement was less than 6% when the training sample size increased from a half to a full training dataset (ie, twice). Training duration on a full training dataset was 74.4 hours. The segmentation time was 15 seconds per slice.

Figure 4 additionally demonstrates the performance of state-of-the-art networks on a development dataset in comparison with the PB-CNN proposed here. For the classical image-based U-Net CNN, DSC was much lower (0.64). The segmentation time was 0.75 seconds per slice. For the PB variant of the U-Net CNN, the training time per epoch was 23 times longer than for our PB-CNN, which deemed it infeasible to train the network in an acceptable time. Therefore, the network was trained using a reduced training patch database (~1/20 of the full one) to stay within the feasibility limit. The corresponding DSC value was 0.44 and the segmentation time was 2.05 minutes per slice.

### 3.2.2 | PB-CNN performance validation

Results for the CNN-based cartilage segmentation procedure accuracy for the method validation phase are summarized in Table 1. The averaged DSC was  $0.81 \pm 0.05$ . The averaged cartilage CSA ( $266.5 \pm 34.3 \text{ mm}^3$ ) was significantly higher ( $P = .019$ ) than the corresponding value quantified by the expert ( $237.6 \pm 39.8 \text{ mm}^3$ ). According to the paired t-test ( $P > .05$ ), the segmentation accuracy was not influenced by the coil selection.

Examples of CNN-based segmentations of different accuracies are displayed in Figure 5. The CNN-based segmentation of the validation dataset was qualitatively assessed by an independent radiologist who scored  $7.00 \pm 1.51$  for controls and  $4.60 \pm 1.14$  for patients (Table 1). The scores were consistent with the average DSC values for these cartilage masks ( $0.82 \pm 0.03$  and  $0.76 \pm 0.04$  for healthy volunteers and patients, respectively).

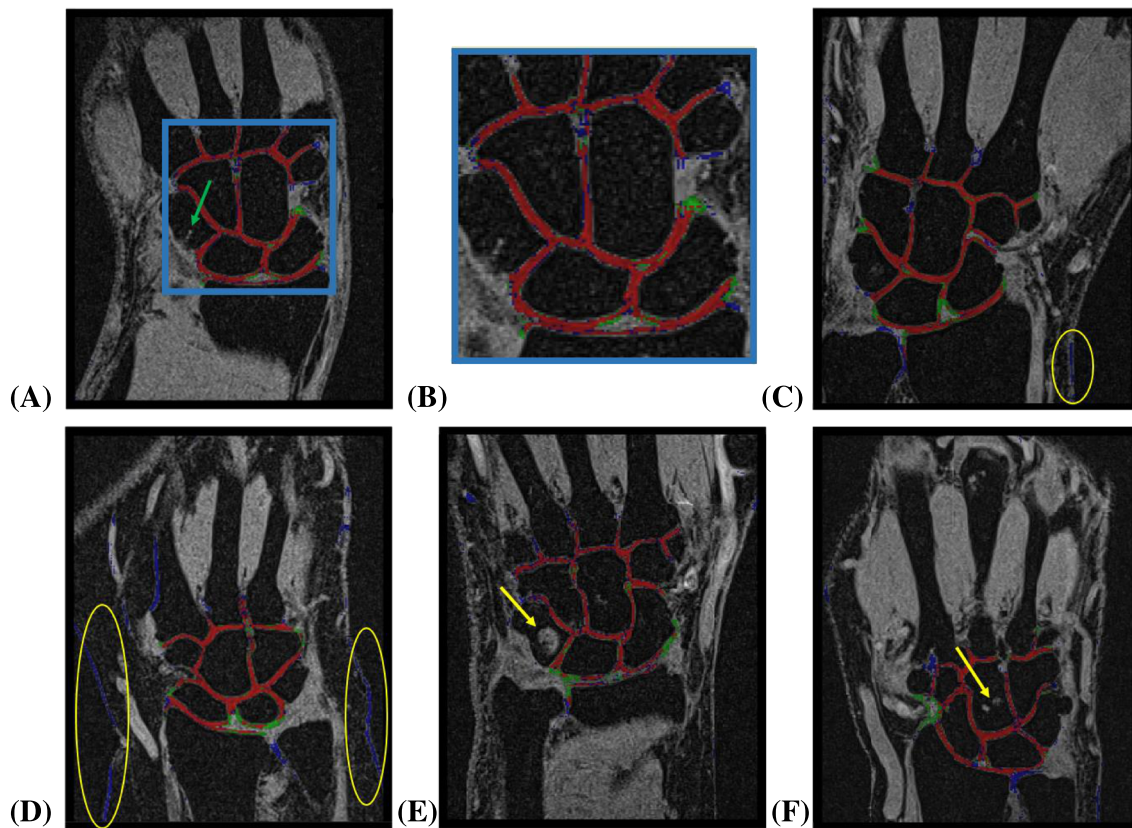
### 3.2.3 | Performance of PB-CNN across 3D volume

Table 2 shows DSC scores for layer-to-layer experiments, which involved analysis of different plane locations in 3D. DSC values averaged across the 3D images of the testing dataset were  $0.69 \pm 0.06$  for the whole group,  $0.73 \pm 0.03$  for the healthy subjects, and  $0.65 \pm 0.05$  for the patients. The averaged DSC value in the cross-validation analysis was  $0.70 \pm 0.05$  ( $0.73 \pm 0.02$  and  $0.67 \pm 0.05$  for controls and patients, respectively). The layer-based analysis showed that the DSC was lower for the slices located far away from the medial cross-section, both for “hold-out” and cross-validation (in brackets) tests: zone#1:  $0.21 \pm 0.21$  ( $0.25 \pm 0.20$ ), zone#2:  $0.60 \pm 0.09$  ( $0.61 \pm 0.09$ ), zone#3:  $0.63 \pm 0.06$  ( $0.65 \pm 0.05$ ), and zone#4:  $0.73 \pm 0.05$  ( $0.74 \pm 0.05$ ).

## 4 | DISCUSSION

The aim of the present study was to investigate the performance of a dedicated CNN for the segmentation of the wrist cartilage from structural MR images, to compare the corresponding results with manual and existing CNN-based segmentation approaches, and to assess the dependence of the network’s performance on the amount and heterogeneity of the training data.

Our results demonstrated that the presented PB-CNN architecture significantly outperformed the classical image-based U-Net in the wrist cartilage segmentation task (DSC = 0.86 and 0.64, respectively). This improvement came at the expense of the computational time (15 seconds



**FIGURE 5** Illustrations of performance of the proposed PB-CNN (red: correctly segmented pixels [true positives]; green: pixels incorrectly assigned to the background [false negatives]; and blue: pixels incorrectly assigned to the cartilage [false positives]). (A) Representative segmentation example (healthy subject, medial slice, DSC = 0.86, visual evaluation score = 8); zoomed-in cartilage area is shown in (B). The green arrow points to a vessel that had contrast and geometry similar to cartilage but was not assigned to this type of tissue by our PB-CNN. (C) Additional segmentation example (healthy subject, medial slice, DSC = 0.81, visual evaluation score = 6). (D) Example of segmentation of patient data with diminished performance (medial slice, DSC = 0.69, visual evaluation score = 3). The yellow circles show the skin tissue considered by CNN as cartilage. (E, F) Additional illustrations of CNN performance on the images of patients. The yellow arrows point to the high signal intensity lesions, which were correctly excluded by the proposed PB-CNN from the segmented mask

**TABLE 2** Results of layer-to-layer analysis of CNN segmentation performance: DSC averaged over zones for “hold-out” and cross-validation (in brackets) studies

Subjects group	No cartilage zone#1	1%-33% zone#2	34%-66% zone#3	67%-100% zone#4	3D DSC
Averaged over HV group ± SD	0.24 (0.28) ± 0.20 (0.20)	0.66 (0.66) ± 0.04 (0.04)	0.67 (0.68) ± 0.05 (0.05)	0.76 (0.77) ± 0.03 (0.03)	0.73 (0.73) ± 0.03 (0.02)
Averaged over P group ± SD	0.18 (0.22) ± 0.24 (0.21)	0.54 (0.56) ± 0.08 (0.11)	0.58 (0.61) ± 0.03 (0.03)	0.69 (0.72) ± 0.05 (0.05)	0.65 (0.67) ± 0.05 (0.05)
Averaged over all scans ± SD	0.21 (0.25) ± 0.21 (0.20)	0.60 (0.61) ± 0.09 (0.09)	0.63 (0.65) ± 0.06 (0.05)	0.73 (0.74) ± 0.05 (0.05)	0.69 (0.70) ± 0.06 (0.05)

Abbreviations: CNN, convolutional neural network; DSC, Sørensen–Dice similarity coefficient; HV, healthy volunteers (MRI scans #11–#15); P, patients (MRI scans #16–#20); SD, standard deviation

for our PB-CNN vs. 0.75 seconds for the image-based U-Net CNN). The decreased accuracy of the image-based network may be explained by the relatively low number of training samples, which, however, did not result in reduced performance of our PB method. Our PB-CNN also significantly outperformed the PB-U-Net architecture under identical training conditions (Figure 4). Interestingly, our network was much faster than PB-U-Net, which may be explained by a much lower number of parameters in our planar PB-CNN and the need for a full patch mask for PB-U-Net training. It should be noted that we utilized basic U-net architectures that were not fine-tuned for the wrist joint cartilage segmentation on MR images. Overall, our PB-CNN provided fast and reliable segmentation of wrist cartilage in MR images.

The performance of our PB-CNN agreed well with that of the manual segmentation, as evidenced by the comparable DSC values ( $0.81 \pm 0.05$ ) with those from the inter-observer assessment ( $0.88 \pm 0.04$  for O1 vs. O2 and  $0.78 \pm 0.06$  for O1 vs. O3 in medial slices). At the



same time, it was lower compared with the intra-observer study ( $0.90 \pm 0.04$ ), thereby indicating that the PB-CNN did not fully reproduce the manual segmentation strategy of O1 (one must keep in mind that the same observer [O1] segmented the wrist cartilage for training and for the intra-observer study). However, the DSC values were comparable with those previously reported for CNN-assisted knee cartilage segmentation ( $0.82\text{--}0.88$ ),<sup>25,26</sup> which demonstrates the potential of CNN-based methods for more challenging wrist cartilage segmentation.

Our network demonstrated robustness regarding many anatomical structures and joint abnormalities that have appearances or contrast similar to cartilage, as illustrated by representative examples in Figure 5 (eg, a vessel in the capitate bone of a healthy volunteer (Figure 5A) and cyst and cortical bone erosions in patients (Figure 5E,F). At the same time, the analysis indicated an elevated number of false positive pixels in images of patients, mostly due to misclassification of skin tissue as cartilage (Figure 5D). Our cross-validation study demonstrated that the PB-CNN segmentation accuracy of patients' data benefited from larger heterogeneity of the training dataset. Further, despite the fact that relatively few subjects and MRI scans were included in our study, the segmentation accuracy growth saturated quickly with increasing size of the training dataset (Figure 4). This suggests that the biological variability may be a more important characteristic of the training dataset compared with its size. Hence, clinical implementation of the technique may require additional training of the network on a more heterogeneous dataset. The enhanced training would benefit from including patients with varying age and body mass, the biological factors contributing to structural and image contrast variabilities. Further, the method's performance in clinical cases may potentially be improved by explicit inclusion of lesions and anatomical structures other than cartilage into the labeled training database as previously described.<sup>27</sup>

The current study considered computer-assisted manual segmentation as a ground truth for dataset labeling purposes. Our results demonstrate that the implemented procedure was sufficiently reproducible, with inter-observer DSC reaching  $0.88 \pm 0.04$  (O1 and O2). This performance is similar to the inter-observer DSC reported for the manual segmentation of knee cartilage ( $0.88^{36}$ ), which is less challenging to segment than cartilage in wrist due to the higher thickness and anatomical complexity of the former. However, the agreement between O1 and O3 was somewhat lower (DSC =  $0.78 \pm 0.06$ ). The discordance may be partially explained by differences in training and inter-institutional differences (O3 was affiliated to a different institution than O1 and O2). At the same time, the observation suggests that merging multi-institutional labeled datasets into a single training dataset could be an appropriate training strategy to avoid the bias caused by the segmentation practices adapted by each individual observer and/or research site.

Our study demonstrated a heterogeneous performance of the segmentation with respect to the slice location (Table 2). The best performance was achieved for medial cross-sections of the wrist, in which cartilage tissue is characterized by the most well-defined and interconnected geometry. The worst performance was observed in the slices located away from the medial cross-section, in which the amount of cartilage is low, and may become less recognizable by the network due to poorly defined morphological features. Considering that the cross-validation did not significantly improve the segmentation quality at the periphery relative to the medial zone (Table 2), we suggest that increasing the number of subjects included in the training dataset may not be the most efficient strategy to improve performance in those locations. At the same time, it is worth noting that increasing the number of slices chosen from each 3D image was not studied separately for lateral slices and should be a subject for further investigation. The major improvement may come from exploiting 3D peculiarities of cartilage, which are different compared with skin or vessels. Exploiting such spatial correlations would require expanding the network architecture into 3D.<sup>37</sup> However, the challenges of such modification are related to a very high computation and memory cost of the 3D PB approach<sup>38</sup> and implementation of 3D image labeling.

## 5 | CONCLUSIONS

The PB-CNN-based segmentation of wrist cartilage from MR images proposed here provides a time-efficient alternative to manual segmentation. The proposed architecture outperformed a state-of-the-art image-based U-Net architecture and its PB variant when trained on a limited amount of subjects' data. Our results highlight the importance of including a sufficient number of patients in the training dataset. The accuracy of the proposed approach might be further increased with a more heterogeneous multi-institutional training sample and by using a 3D CNN architecture.

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## REFERENCES

1. Eckstein F, Kunz M, Schutzer M, et al. Two year longitudinal change and test-retest-precision of knee cartilage morphology in a pilot study for the osteoarthritis initiative. *Osteoarthr Cartil.* 2007;15(11):1326-1332.
2. Visser AW, Bøyesen P, Haugen IK, et al. Radiographic scoring methods in hand osteoarthritis – a systematic literature search and descriptive review. *Osteoarthr Cartil.* 2014;22(10):1710-1723.
3. Link T, Stahl R, Woertler K. Cartilage imaging: Motivation, techniques, current and future significance. *Eur Radiol.* 2007;17(5):1135-1146.
4. Link T. *Cartilage Imaging: Significance, Techniques, and New Developments.* New York, NY: Springer Science & Business Media; 2011:93-102.
5. Eckstein F, Kunz M, Schutzer M, et al. Magnetic resonance imaging (MRI) of articular cartilage in knee osteoarthritis (OA): morphological assessment. *Osteoarthr Cartil.* 2006;14:46-75.
6. Ostergaard M, Bøyesen P, Eshed I, et al. Development and preliminary validation of a magnetic resonance imaging joint space narrowing score for use in rheumatoid arthritis: potential adjunct to the OMERACT RA MRI scoring system. *J Rheumatol.* 2011;38(9):2045-2050.
7. Zink JV, Souteyrand P, Guis S, et al. Standardized quantitative measurements of wrist cartilage in healthy humans using 3T magnetic resonance imaging. *World J Orthop.* 2015;6(8):641-648.
8. Peterfy CG, Van Dijke CF, Janzen DL, et al. Quantification of articular cartilage in the knee with pulsed saturation transfer subtraction and fat-suppressed MR imaging: optimization and validation. *Radiology.* 1994;192(2):485-491.
9. Peterfy CG, Van Dijke CF, Lu Y, et al. Genant HK Quantification of the volume of articular cartilage in the metacarpophalangeal joints of the hand: accuracy and precision of three-dimensional MR imaging. *Am J Roentgenol.* 1995;165(2):371-375.
10. Duvvuri U, Kudchodkar S, Reddy R, Leigh JS. T(1rho) relaxation can assess longitudinal proteoglycan loss from articular cartilage in vitro. *Osteoarthr Cartil.* 2002;10:838-844.
11. Ling W, Regatte RR, Navon G, Jerschow A. Assessment of glycosaminoglycan concentration in vivo by chemical exchange-dependent saturation transfer (gagCEST). *Proc Natl Acad Sci U S A.* 2008;105(7):2266-2270.
12. Liu F, Block WF, Kijowski R, Samsonov A. Rapid multi-component relaxometry in steady state with correction of magnetization transfer effects. *Magn Reson Med.* 2016;75:1423-1433.
13. Sritanyaratana N, Samsonov A, Mossahebi P, Wilson JJ, Block WF, Kijowski R. Cross-relaxation imaging of human patellar cartilage in-vivo at 3.0T. *Osteoarthr Cartil.* 2014;22(10):1568-1578.
14. Nishimura K, Tanabe T, Kimura M, Harasawa A, Karita K, Matsushita T. Measurement of articular cartilage volumes in the normal knee by magnetic resonance imaging: can cartilage volumes be estimated from physical characteristics? *J Orthop Sci.* 2005;10(3):246-252.
15. Sittek H, Eckstein F, Gavazzeni A, et al. Assessment of normal patellar cartilage volume and thickness using MRI: an analysis of currently available pulse sequences. *Skeletal Radiol.* 1996;25(1):55-62.
16. Maataoui A, Graichen H, Abolmaali ND, et al. Quantitative cartilage volume measurement using MRI: comparison of different evaluation techniques. *Eur Radiol.* 2005;15(8):1550-1554.
17. Piplani MA, Disler DG, McCauley TR, Holmes TJ, Cousins JP. Articular cartilage volume in the knee: semiautomated determination from three-dimensional reformations of MR images. *Radiology.* 1996;198:855-859.
18. Liukkonen MK, Mononen ME, Tanska P, Sarakkala S, Nieminen MT, Korhonen RK. Application of a semi-automatic cartilage segmentation method for biomechanical modeling of the knee joint. *Comput Methods Biomech Biomed Engin.* 2017;20(13):1453-1463.
19. Fernquest S, Park D, Marcan M, Palmer A, Voiculescu I, Glyn-Jones S. Segmentation of hip cartilage in compositional magnetic resonance imaging: A fast, accurate, reproducible, and clinically viable semi-automated methodology. *J Orthop Res.* 2018;36:2280-2287.
20. Folkesson J, Dam EB, Olsen OF, Petttersen PC, Christiansen C. Segmenting articular cartilage automatically using a voxel classification approach. *IEEE Trans Med Imaging.* 2007;26(1):106-115.
21. Kashyap S, Oguz I, Zhang H, Sonka M. Automated Segmentation of Knee MRI Using Hierarchical Classifiers and Just Enough Interaction Based Learning: Data from Osteoarthritis Initiative. In: *Springer, International Conference on Medical Image Computing and Computer-Assisted Intervention.* Vol.9901 Cham, Switzerland: Springer; 2016:344-351.
22. Vincent G, Wolstenholme C, Scott I, Bowes M. Fully automatic segmentation of the knee joint using active appearance models. In: *Proceedings of MICCAI Workshop on Medical Image Analysis for the Clinic.* Berlin, Germany: Springer; 2010:224-230.
23. Lee JG, Gumus S, Moon CH, Kwok CK, Bae KT. Fully automated segmentation of cartilage from the MR images of knee using a multi-atlas and local structural analysis method. *Med Phys.* 2014;41(9):092303.
24. Dice LR. Measures of the Amount of Ecologic Association Between Species. *Ecology.* 1945;26(3):297-302.
25. Prasoon A, Petersen K, Igel C, Lauze F, Dam E, Nielsen M. Deep feature learning for knee cartilage segmentation using a triplanar convolutional neural network. *Med Image Comput Comput Assist Interv.* 2013;16(Pt 2):246-253.
26. Norman B, Pedoia V, Majumdar S. Use of 2D U-Net convolutional neural networks for automated cartilage and meniscus segmentation of knee MR imaging data to determine relaxometry and morphometry. *Radiology.* 2018;288(1):172-185.
27. Liu F, Zhou Z, Jang H, Samsonov A, Zhao G, Kijowski R. Deep convolutional neural network and 3D deformable approach for tissue segmentation in musculoskeletal magnetic resonance imaging. *Magn Reson Med.* 2018;79:2379-2391.
28. Zhou Z, Zhao G, Kijowski R, Liu F. Deep convolutional neural network for segmentation of knee joint anatomy. *Magn Reson Med.* 2018;80(6):2759-2770.
29. Ronneberger O, Fischer P, Brox T. U-Net: Convolutional Networks for Biomedical Image Segmentation. *Med Image Comput Comput Assist Interv.* 2015;9351:234-241.
30. Li AE, Lee SK, Rancy SK, Burge AJ, Potter HG, Wolfe SW. Comparison of magnetic resonance imaging and radiographs for evaluation of carpal osteoarthritis. *J Wrist Surg.* 2017;6(2):120-125.
31. Moser T, Dosch JC, Moussaoui A, Buy X, Gangi A, Dietemann JL. Multidetector CT arthrography of the wrist joint: how to do it. *Radiographics.* 2008;28:787-800.
32. Sekou TB, Hidane M, Olivier J, Cardot H. From Patch to Image Segmentation using Fully Convolutional Networks - Application to Retinal Images. arXiv:1904.03892v2 [cs.CV].
33. Shchelokova AV, Van den Berg CAT, Dobrykh DA, et al. Volumetric wireless coil based on periodically coupled split-loop resonators for clinical wrist imaging. *Magn Reson Med.* 2018;80(4):1726-1737.

34. Hou L, Samaras D, Kurc TM, Gao Y, Davis JE, Saltz JH. Patch-Based Convolutional Neural Network for Whole Slide Tissue Image Classification. In: 2016 IEEE Conference on Computer Vision and Pattern Recognition (CVPR). Silver Spring, United States: IEEE Computer Society Press; 2016;2424-2433.
35. Hesamian MH, Jia W, He X, Kennedy P. Deep learning techniques for medical image segmentation: achievements and challenges. *J Digit Imaging*. 2019;32:582-596.
36. Shim H, Chang S, Tao C, Wang JH, Kwok CK, Bae KT. Knee cartilage: efficient and reproducible segmentation on high-spatial-resolution MR images with the semiautomated graph-cut algorithm method. *Radiology*. 2009;251(2):548-556.
37. Pezeshk A, Hamidian S, Petrick N, Sahiner B. 3D convolutional neural networks for automatic detection of pulmonary nodules in chest CT. *IEEE J Biomed Health Inform*. 2019;23(5):2080-2090.
38. Dou Q, Chen H, Yu L, et al. Automatic detection of cerebral microbleeds from MR images via 3D convolutional neural networks. *IEEE Trans Med Imaging*. 2016;35(5):1182-1195.

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## APPENDIX A

**FIGURE A1** Summary of the PB-CNN architecture optimized for wrist cartilage segmentation

Layer (type)	Output Shape	Param #
gaussian_noise_2 (GaussianNo	(None, 28, 28, 1)	0
alpha_dropout_2 (AlphaDropou	(None, 28, 28, 1)	0
conv2d_6 (Conv2D)	(None, 26, 26, 44)	440
max_pooling2d_2 (MaxPooling2	(None, 13, 13, 44)	0
conv2d_7 (Conv2D)	(None, 11, 11, 44)	17468
conv2d_8 (Conv2D)	(None, 9, 9, 44)	17468
conv2d_9 (Conv2D)	(None, 7, 7, 44)	17468
conv2d_10 (Conv2D)	(None, 5, 5, 44)	17468
flatten_2 (Flatten)	(None, 1100)	0
dense_2 (Dense)	(None, 2)	2202
=====		
Total params: 72,514		
Trainable params: 72,514		
Non-trainable params: 0		

Layer (type)	Output Shape	Param #	Connected to
input_5 (InputLayer)	(None, 272, 256, 1)	0	
lambda_5 (Lambda)	(None, 272, 256, 1)	0	input_5[0][0]
conv2d_73 (Conv2D)	(None, 272, 256, 64)	128	lambda_5[0][0]
conv2d_74 (Conv2D)	(None, 272, 256, 64)	4160	conv2d_73[0][0]
max_pooling2d_17 (MaxPooling2D)	(None, 136, 128, 64)	0	conv2d_74[0][0]
conv2d_75 (Conv2D)	(None, 136, 128, 128)	8320	max_pooling2d_17[0][0]
conv2d_76 (Conv2D)	(None, 136, 128, 128)	16512	conv2d_75[0][0]
max_pooling2d_18 (MaxPooling2D)	(None, 68, 64, 128)	0	conv2d_76[0][0]
conv2d_77 (Conv2D)	(None, 68, 64, 256)	33024	max_pooling2d_18[0][0]
conv2d_78 (Conv2D)	(None, 68, 64, 256)	65792	conv2d_77[0][0]
max_pooling2d_19 (MaxPooling2D)	(None, 34, 32, 256)	0	conv2d_78[0][0]
conv2d_79 (Conv2D)	(None, 34, 32, 512)	131584	max_pooling2d_19[0][0]
conv2d_80 (Conv2D)	(None, 34, 32, 512)	262656	conv2d_79[0][0]
max_pooling2d_20 (MaxPooling2D)	(None, 17, 16, 512)	0	conv2d_80[0][0]
conv2d_81 (Conv2D)	(None, 17, 16, 1014)	520182	max_pooling2d_20[0][0]
conv2d_transpose_17 (Conv2DTran	(None, 34, 32, 512)	2077184	conv2d_81[0][0]
concatenate_17 (Concatenate)	(None, 34, 32, 1024)	0	conv2d_transpose_17[0][0] conv2d_80[0][0]
conv2d_82 (Conv2D)	(None, 34, 32, 512)	524800	concatenate_17[0][0]
conv2d_83 (Conv2D)	(None, 34, 32, 512)	262656	conv2d_82[0][0]
conv2d_transpose_18 (Conv2DTran	(None, 68, 64, 256)	524544	conv2d_83[0][0]
concatenate_18 (Concatenate)	(None, 68, 64, 512)	0	conv2d_transpose_18[0][0] conv2d_78[0][0]
conv2d_84 (Conv2D)	(None, 68, 64, 256)	131328	concatenate_18[0][0]
conv2d_85 (Conv2D)	(None, 68, 64, 256)	65792	conv2d_84[0][0]
conv2d_transpose_19 (Conv2DTran	(None, 136, 128, 128)	131200	conv2d_85[0][0]
concatenate_19 (Concatenate)	(None, 136, 128, 256)	0	conv2d_transpose_19[0][0] conv2d_76[0][0]
conv2d_86 (Conv2D)	(None, 136, 128, 128)	32896	concatenate_19[0][0]
conv2d_87 (Conv2D)	(None, 136, 128, 128)	16512	conv2d_86[0][0]
conv2d_transpose_20 (Conv2DTran	(None, 272, 256, 64)	32832	conv2d_87[0][0]
concatenate_20 (Concatenate)	(None, 272, 256, 128)	0	conv2d_transpose_20[0][0] conv2d_74[0][0]
conv2d_88 (Conv2D)	(None, 272, 256, 64)	8256	concatenate_20[0][0]
conv2d_89 (Conv2D)	(None, 272, 256, 64)	4160	conv2d_88[0][0]
conv2d_90 (Conv2D)	(None, 272, 256, 1)	65	conv2d_89[0][0]
Total params: 4,854,583			
Trainable params: 4,854,583			
Non-trainable params: 0			

**FIGURE A2** Summary of the basic image-based-U-Net CNN architecture considered as the state-of-the-art architecture for medical image segmentations

Layer (type)	Output Shape	Param #	Connected to
input_1 (InputLayer)	(None, 28, 28, 1)	0	
lambda_1 (Lambda)	(None, 28, 28, 1)	0	input_1[0][0]
conv2d_1 (Conv2D)	(None, 28, 28, 64)	128	lambda_1[0][0]
conv2d_2 (Conv2D)	(None, 28, 28, 64)	4160	conv2d_1[0][0]
max_pooling2d_1 (MaxPooling2D)	(None, 14, 14, 64)	0	conv2d_2[0][0]
conv2d_3 (Conv2D)	(None, 14, 14, 128)	8320	max_pooling2d_1[0][0]
conv2d_4 (Conv2D)	(None, 14, 14, 128)	16512	conv2d_3[0][0]
max_pooling2d_2 (MaxPooling2D)	(None, 7, 7, 128)	0	conv2d_4[0][0]
conv2d_5 (Conv2D)	(None, 7, 7, 256)	33024	max_pooling2d_2[0][0]
conv2d_transpose_1 (Conv2DTrans	(None, 14, 14, 128)	131200	conv2d_5[0][0]
concatenate_1 (Concatenate)	(None, 14, 14, 256)	0	conv2d_transpose_1[0][0] conv2d_4[0][0]
conv2d_6 (Conv2D)	(None, 14, 14, 128)	32896	concatenate_1[0][0]
conv2d_7 (Conv2D)	(None, 14, 14, 128)	16512	conv2d_6[0][0]
conv2d_transpose_2 (Conv2DTrans	(None, 28, 28, 64)	32832	conv2d_7[0][0]
concatenate_2 (Concatenate)	(None, 28, 28, 128)	0	conv2d_transpose_2[0][0] conv2d_2[0][0]
conv2d_8 (Conv2D)	(None, 28, 28, 64)	8256	concatenate_2[0][0]
conv2d_9 (Conv2D)	(None, 28, 28, 64)	4160	conv2d_8[0][0]
conv2d_10 (Conv2D)	(None, 28, 28, 1)	65	conv2d_9[0][0]
Total params: 288,065			
Trainable params: 288,065			
Non-trainable params: 0			

**FIGURE A3** Summary of the basic patch-based-U-Net CNN architecture considered as the state-of-the-art patch-based architecture for medical image segmentations